CELL EXPANSION GENES EXPRESSION BY THERAPEUTIC ULTRASOUND. PROS AND CONS.

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1. INTRODUCTION

Mesenchymal stem cells (MSCs) have the capabilities for self-renewal and differentiation into cells with the phenotypes of, among others, bone, cartilage, and fat. Because bone marrow derived cells, which are a main source of MSCs, are not always acceptable due to a significant drop in their cell number and proliferative/differentiation capacity with age, there is a need for a technique that is capable of expansion of these cells or finding an alternative stem cell source.

Nucleostemin is a newly discovered nucleolar protein present in both embryonic and adult rat central nervous system stem cells, and several human cancer cell lines [1]. It has been reported also that Nucleostemin gene is a marker of proliferating stromal stem cells in adult human bone marrow [2].

Ultrasound, an acoustic pressure wave at frequencies above the limit of human hearing, is transmitted into and through biological tissues and is being used widely in medicine as a therapeutic, operative, and diagnostic tool.[3, 4]. It has been reported that Low Intensity Pulsed Ultrasound (LIPUS) enhances different types of cell proliferation that includes chondrocytes [5]; skin fibroblasts [6] and increase VEGF vascular endothelial growth factor (VEGF), a growth factor associated with endothelial cell proliferation and migration [7]. The effect of LIPUS on stem cells is not fully studied or understood. It also has been shown that ultrasound stimulates tendon cell proliferation and upregulate of proliferating cell nuclear antigen (PCNA) [8]. Also, therapeutic ultrasound upregulate c-myc, a proto-oncogene, binds DNA through its basic helix-loop-helix/leucine zipper domain, which is essential to cell proliferation and differentiation [9].

Human Umbilical Cord PeriVascular (HUCPV) cells are a potential substitute for BMCs due to the immaturity of newborn cells [10]. Umbilical cord blood transplantation (UCBT) has become an established haematopoietic stem cell therapy for patients with no reported immune-rejection. [10]. Due to donor number limitation is a major constraint to bone marrow mesenchymal stem cell therapy, there is a high need for alternative cell sources for such cell-based therapies [11].

Taking the aforementioned information, we hypothesized that LIPUS can up-regulate the following genes Nucleostemin gene in rapidly proliferative and self renewal cells (Bone marrow stem cells, Human Umbilical perivascular endothelial cells...

2. METHOD

2.1 Cells

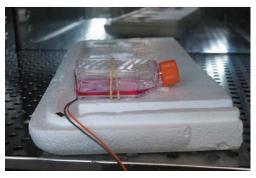
Bone marrow stem cells (BMSCs) were isolated from tibiae and femora of -3 month old male Sprague-Dawley rats (200gms). The femora will be extracted, and the bone marrow cells were flushed with basic media using 10-ml syringe from both sides. Then the cells were centrifuged for 6 min at 600 rpms and were resuspend in tissue culture media, and plated at 5 x 107 cell/mm culture dish (6-well plate), then were incubated in 95% and CO2 at 37° C, with fresh media every 3 days. Cells were expanded and trypsenized and passaged after 2 weeks. Cells were seeded in 75 mm flasks. Cells were expanded in culture medium that contained (Dulbecco's Modified Eagle's Medmium-low Glucose medium (DMEM-LG), 10% fetal bovine serum (FBS), 1% streptomycin/penicillin, and ascorbic acid (50 µg/ml).

Human Umbilical Cord PeriVascular (HUCPV) were donated by Dr. JE Davis, UT, Toronto, ON, Canada. Details about isolation and behavior of these cells are reported previously [11].

2.2 LIPUS

LIPUS was applied for twenty minutes per day to the base of the cell culture flasks. LIPUS was applied using a 2.5 cm2 transducer that delivers 1.5 MHz with a duty cycle of ¼ and delivers SATA intensity of 30 mW/cm2. The power output was calibrated weekly to ensure that the output is maintained throughout the experiment. Ultrasound was applied to the bottom of each cell flask for 20 minutes per day for 7 and 20 days (Figure 1).

Fig. 1. Application of LIPUS to the cell culture flasks.



2.3 RT-PCR

Primers for the Nucleostemin were as follows: Forward:TCCGAAGTCCAGCAAGTATTG; and reverse: AATGAGGCACCTGTCCACTC. The housekeeping gene glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as a control. Samples were amplified for 22 - 30 cycles in an Eppendorf Master-cycler thermal cycler. PCR reactions were electrophoresed on 2% agarose gels and

visualized under UV light after staining with Ethidium Bromide using image analyzer.

3. RESULTS

3.1 Cell response to LIPUS.

Figures 1 and 2 show PCR reactions gel electrophoresis. It appears that LIPUS has increased Nucleostemin expression in both types of stem cells.

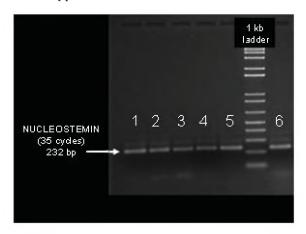


Fig. 1. Nucletemin up-regulation by LIPUS after 7 days in rat BMSCs. RT-PCR of mRNA from BMCs in Culture. 1,2 (T0), 3,4 (Control), 5,6 (LIPUS).

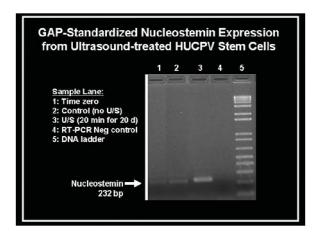


Fig. 2. Nucletemin up-regulation by LIPUS after 20 days (3) compared to control (2) lanes in HUCPVSCs.

4. DISCUSSION

Our results support previous findings that LIPUS can increase gene expression by different cells. The increased gene expression of Nucleostemin is in accordance with increased cell proliferation. Rat bone marrow stem cells responded positively to LIPUS by increased Nucleostemin expression after 7 days, while HUCPVSCs responded to LIPUS by increased expression of Nucleostemin expression by LIPUS after 20 days. This preliminary study suggests that optimization experiment to test expression Nucleostemin by each type of stem cells be conducted.

Our data suggests that Therapeutic ultrasound might be beneficial in enhancing proliferation of stem cells and maintaining their pluripotent characteristic by continuing expression of Nucleostemin. However this might be a negative effect on neoplastic cells. Since Nucleostemin is also a marker of cancer cells [1], until a study is conducted on the effect of LIPUS on cancer cell proliferation and associated gene expression, it should be recommended that caution must be taken before application of therapeutic application should cancerous cells are expected to be in the vicinity of the application area until further investigation might shows it is safe in such a condition.

REFERENCES

[1] Tsai RY, McKay RD. A (2002) Nucleolar mechanism controlling cell proliferation in stem cells and cancer cells. Genes Dev;16:2991-3003

[2] WAEL KAFIENAH, SANJAY MISTRY, CHRISTOPHER WILLIAMS, ANTHONY P. HOLLANDER (2006). Nucleostemin Is a Marker of Proliferating Stromal Stem Cells in Adult Human Bone Marrow. STEMCELLS;24:1113-1120.

[3] Ziskin, M., Applications of ultrasound in medicinecomparison with other modalities, in Ultrasound: medical applications, biological effects, and hazard potential, M.H. Repacholi, M. Grandolfo, and A. Rindi, Editors. 1987, Plenum

Press: New York. p. 49-59.
[4] Dyson, M., Therapeutic applications of ultrasound., in Biological effects of ultrasound, W.L. Nyborg and M.C. Ziskin,

Editors. 1985, Churchill Livingstone: New York. p. 121-133.

[5] Mukai S, Ito H, Nakagawa Y, Akiyama H, Miyamoto M, Nakamura T.: Transforming growth factor-betal mediates the of effects low-intensity pulsed ultrasound chondrocytes..Ultrasound in Medicine and Biology, 2005. 31(12): p. 1713-1721.

[6] Doan, N., et al., In vitro effects of therapeutic ultrasound on cell proliferation, protein synthesis, and cytokine production by human fibroblasts, osteoblasts, and monocytes. J Oral Maxillofac Surg, 1999. 57(4): p. 409-19; discussion 420.

[7] Reher, P., et al., Effect of ultrasound on the production of IL-

8, basic FGF and VEGF. Cytokine, 1999. 11(6): p. 416-23.
[8] Tsai WC, Hsu CC, Tang FT, Chou SW, Chen YJ, Pang JH.Ultrasound stimulation of tendon cell proliferation and upregulation of proliferating cell nuclear antigen. J Orthop Res. 2005 Jul;23(4):970-6. Epub 2005 Jan 26.PMID: 16023014.

[9] Park, K. S., Lee, Y. S., & Kang, K. S. (2006). In vitro neuronal and osteogenic differentiation of mesenchymal stem cells from human umbilical cord blood. Journal of Veterinary Science, 7(4), 343-348.

[10] Hofmeister, C. C., Zhang, J., Knight, K. L., Le, P., & Stiff, P. J. (2007). Ex vivo expansion of umbilical cord blood stem cells for transplantation: Growing knowledge from the hematopoietic niche. Bone marrow transplantation, 39(1), 11-23.

[11] Sarugaser, R., Lickorish, D., Baksh, D., Hosseini, M. M., & Davies, J. E. (2005). Human umbilical cord perivascular (HUCPV) cells: A source of mesenchymal progenitors. Stem cells, 23(2), 220-229.

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